CELENTYX

IN VITRO SYSTEMS USING HUMAN PRIMARY CELLS AND TISSUES FOR DRUG DEVELOPMENT

IMAGING SLIDESET



www.celentyx.com

COMPANY BACKGROUND; CELENTYX LTD – A SPECIALIST CRO

Founded in 2007 by Professors John Gordon and Nicholas Barnes Provide translational human primary cell and tissue assays for drug development

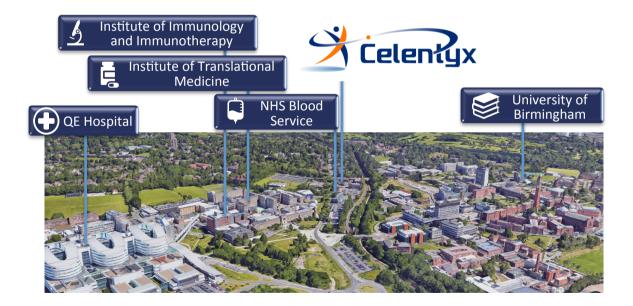


Professor John Gordon



Professor Nicholas Barnes

Applied neuropharmacologist with a proven track record in the investigation of potential drug targets and the delivery of candidate therapeutics







COMPANY BACKGROUND II

Established links to hospitals and clinicians within the Midlands area of the UK

- QE Hospital is the largest single site hospital in UK
- Largest critical care in • Europe
- Serving a large and diverse • community covering a large range of ethnic groups







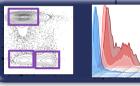
Collection of cells, tissues and/or fluids from patients and 'controls' with subsequent processing for client service projects



All material is consented, collected and stored according to statutory codes of practice



Broad scientific and technical expertise applied across the drug discovery pipeline



Multiparameter flow-cytometry Immune subsets, activation markers, proliferation, target expression



Luminex analysis of fluids

Cytokines, chemokines, other soluble factors



High-throughput confocal microscopy

Plate based high-content screening with automated analysis, tissue immunofluorescence, IHC, RNAScope



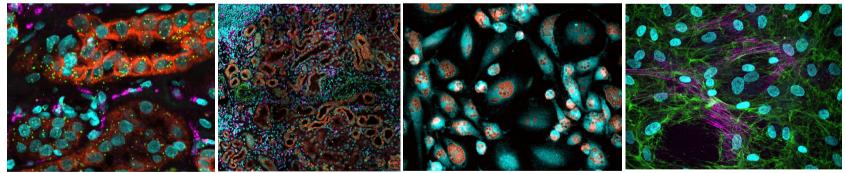
Broad range of other assay techniques available e.g. ELISAs, Western-blotting, calcium imaging, luminescence/ fluorescence assays, radioligand binding



IMAGING ASSAYS

Celentyx Imaging Platform

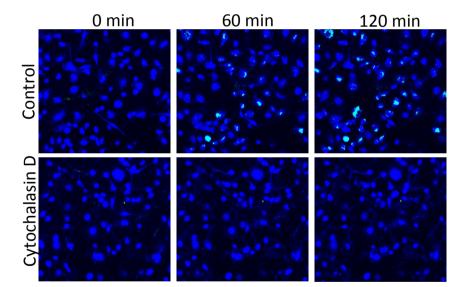
- Range of cutting-edge microscopy platforms available
- Assays can be performed in 96- or 384-well format
- Kinetic, live cell readouts available (including high-speed imaging)
- Multi-colour (4-colour imaging performed routinely; increased colours available)
- Labelling using antibodies, dyes, probes and reporters
- Automated image analysis performed in 2D or 3D
- Applied to a broad range of tissues and cells from patients and healthy donors



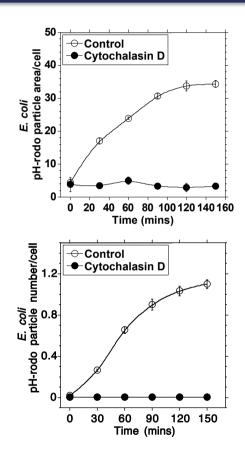


PHAGOCYTOSIS ASSAYS: E. COLI

- Human monocyte-derived macrophages phagocytosing pH-rodo labelled *E.coli*
- Assay performed with or without opsonization of target particle
- Phagocytosis imaged over time by high-throughput spinning disk confocal microscopy
- Time-dependent uptake of *E.coli* by macrophages that is abolished in the presence of cytochalasin D

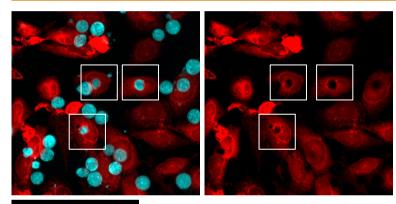






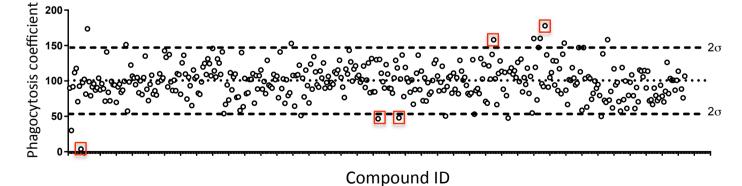
Screening for Phagocytosis Modulators

- High content-imaging screen of a small molecule library for modulators of tumour cell phagocytosis
- Identification of novel modulators that were subsequently validated in orthogonal assays



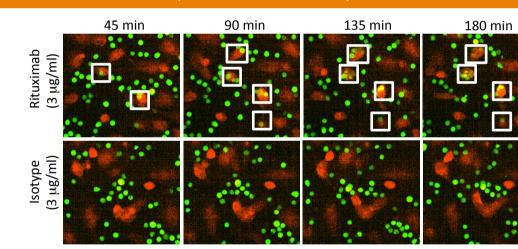
Example screen results shown below. Phagocytosis co-efficient indicates the extent of phagocytosis. Red boxes highlight compounds that were subsequently tested and validated in orthogonal assays.

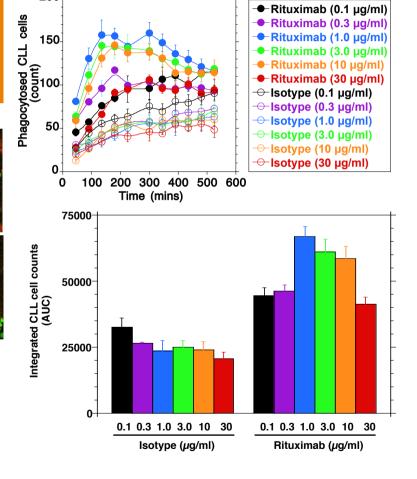
Phagocytic cell Target cell



PHAGOCYTOSIS ASSAYS: TUMOUR CELLS

- Phagocytosis of patient CLL cells by human monocyte-derived macrophages
- Rituximab enhances capture of CLL cells
- Identifies bell-shaped concentration-response



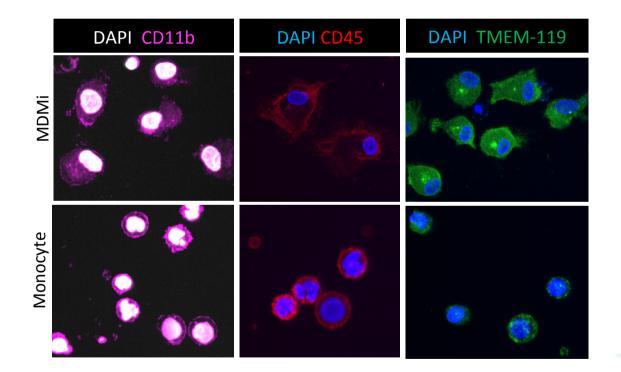


200



INDUCED MICROGLIA FROM MONOCYTES

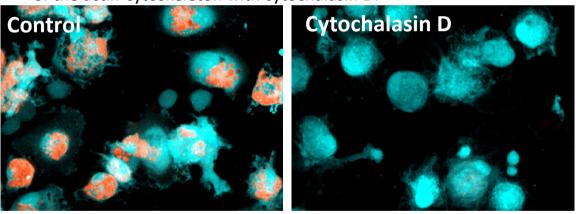
• Human peripheral blood monocytes cultured with a CNS cytokine cocktail to induce differentiation to a microglial phenotype





PHAGOCYTOSIS OF A MYELIN BASIC PROTEIN MEMBRANE PREPARATION

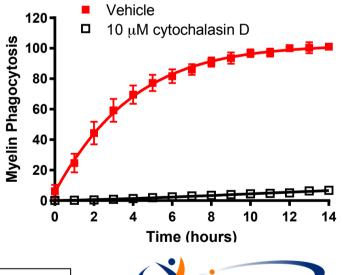
- The phagocytosis of myelin is important during neuronal restructuring and the persistence of myelin debris may contribute to neuroinflammation in a range of diseases
- We have generated human pH-rodo-labelled myelin basic protein (pH-rodo-MBP) for use as a model cargo in phagocytosis assays
- Uptake monitored over time by high-content confocal microscopy in a 96-well plate format
- Time-dependent uptake of pH-rodo-MBP by human monocyte-derived microglia that was sensitive to disruption of the actin-cytoskeleton with cytochalasin D.
 Vehicle





pH-rodo-MBP uptake after 12 hours

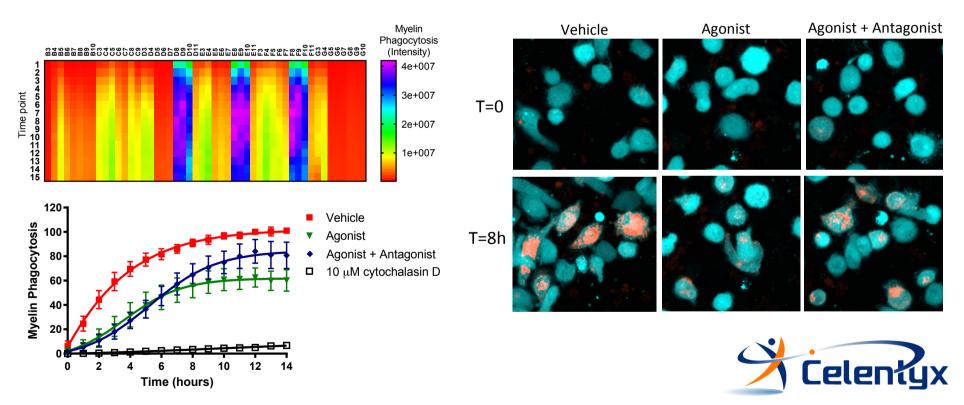
Video of myelin phagocytosis assay available at: <u>http://celentyx.com/imaging_cro_induced_microglia_phagocytosis_assays.html</u>



Celentux

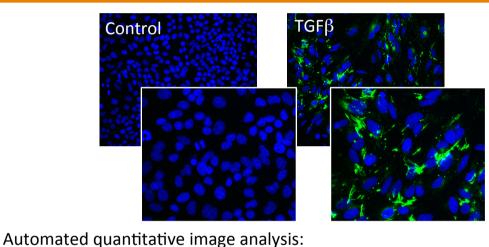
MODULATION OF MYELIN BASIC PROTEIN UPTAKE BY SMALL MOLECULES

Plate-based format allows testing the impact of multiple modulators of phagocytosis



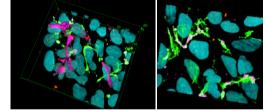
Cell-Based Readouts I

Quantification of fibrotic end-points by high-content imaging



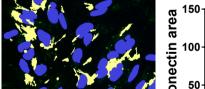
- A human lung epithelial cell line stimulated with TGF β and nuclei and fibronectin stained.
- Confocal Z-stacks from multiple fields of view from 96- or 384well plate acquired and analysed.
- Thickness and overlap between matrix components can also be analysed (Images below).

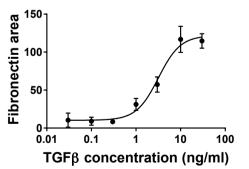
3D projection

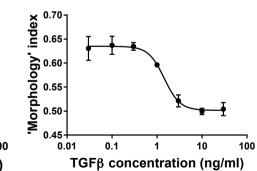








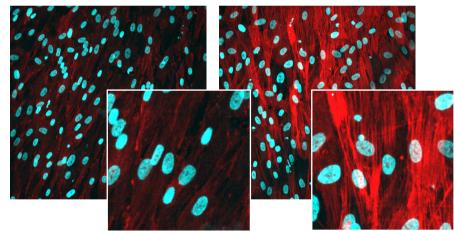


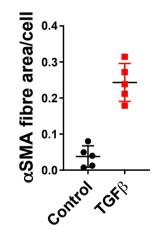


Cell-Based Readouts II

Quantification of α -smooth muscle actin labelling by high-content imaging

- Human primary corneal fibroblasts stimulated with TGF β and nuclei and $\alpha\text{-smooth}$ muscle actin stained
- Confocal Z-stacks from multiple fields of view from 96- or 384-well plate acquired and analysed. Control $$TGF\beta$$

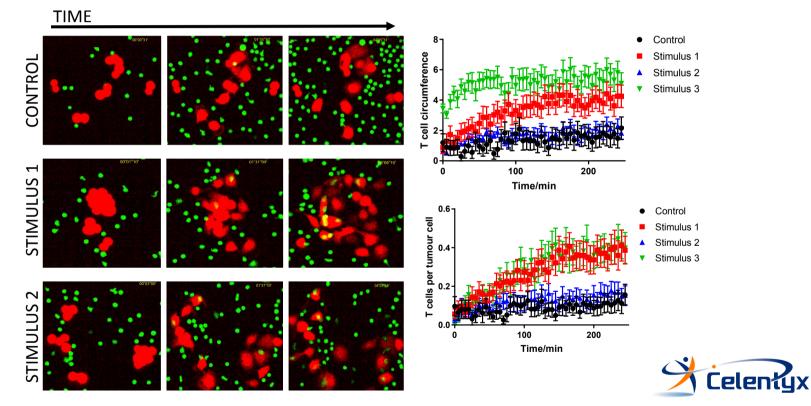






ENGAGEMENT OF TARGET CELLS BY T CELLS

Labeled **human CD8+ T cells** were incubated with **a labelled cell line** Z-stacks were acquired by spinning-disk confocal microscopy

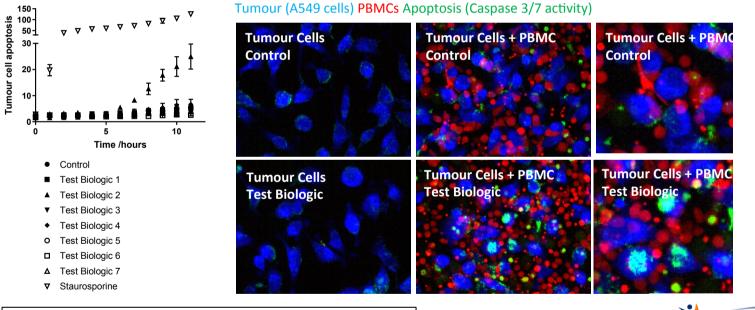


KILLING OF TUMOUR CELLS BY IMMUNE CELLS

Labeled human **PBMC** were incubated with human labelled **tumour cells** Apoptosis measured by **caspase activity**

Z-stacks were acquired by spinning-disk confocal microscopy

Time-dependent killing of tumour cells by PBMC promoted by the test biologic

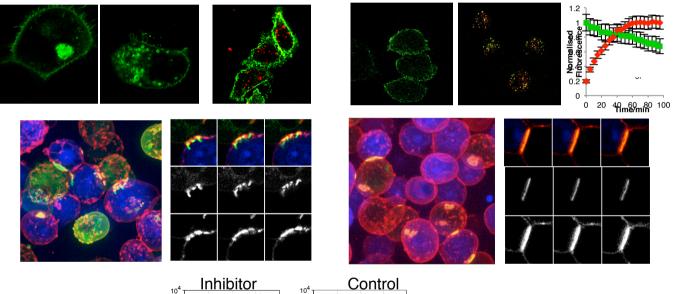


Video of immune cell killing of tumour cells available at: <u>http://celentyx.com/imaging_cro_t_cell_killing.html</u>

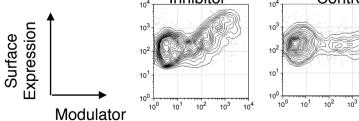


RECEPTOR TRAFFICKING

A range of fluorescence microscopy techniques applied to track and quantify the movement of therapeutics or receptors within cells



104

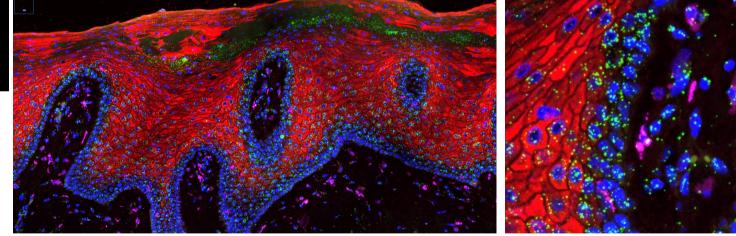




VALIDATION OF TARGET EXPRESSION IN PATIENT TISSUE

- Detection of either target protein by immunofluorescence or when suitable antibodies not available, mRNA-based detection methods such as RNAScope
- Access to comprehensive human tissue bank covering a broad range of indications with associated clinical information - offering investigation of comparable pathological specimens from hundreds of patients for target validation
- Multiplex immunofluorescence analysis (+RNAScope) by confocal microscopy

Tissue, immune cell and target mRNA (RNAScope detection) in patient tissue





SUMMARY

- Celentyx a responsive and knowledgeable CRO, with highest level scientific and technical expertise applied across the drug discovery pipeline including target validation, screening (both primary screens and secondary assays), benchmarking/differentiation studies, safety and mechanism of activation studies
- The internationally recognised leadership team and scientists have a deep knowledge of therapeutic areas involving the immune system including:
 - Immuno-oncology Autoimmunity & inflammation
 - Fibrosis
 - Neuro-inflammation
- An excellent track record designing bespoke research experiments and programmes to provide solutions to client questions
- Extensive links to hospitals and clinicians, providing outstanding access to healthy donor and patient blood and tissue samples (fresh and fixed) with extensive mining of clinical notes
- Utilization of state of the art technologies to deliver high quality research findings
- Flexible and competitive business models from standard fee-for-service to project partnerships
- Contact Dr Catherine Brady at <u>enquiries@celentyx.com</u> to arrange a no obligation discussion with Celentyx's senior scientists to see how the we can help your project



Celentyx Ltd

Birmingham Research park Vincent Drive Edgbaston Birmingham B15 2SQ

T: +44 (0)121 4148199 E: catherine.brady@celentyx.com W: www.celentyx.com



© Celentyx Ltd. 2019